

p. 1

1. a. N-ethylaniline

b. butyl formate

c. 3-iodobutanoic acid

d. 5-methyl-3-hexanone

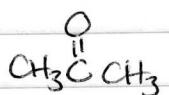
e. 3-chlorobutanal

f. propylamine

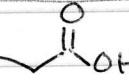
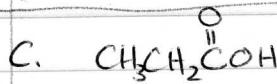
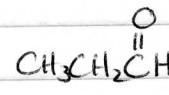
g. N-methyl-N-propylbutanamide

h. N-methylbenzamide

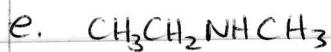
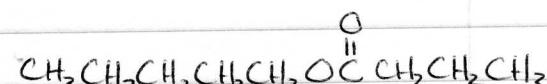
2. a.



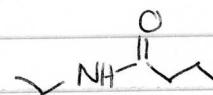
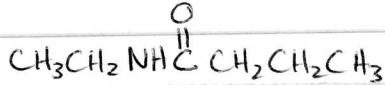
b.



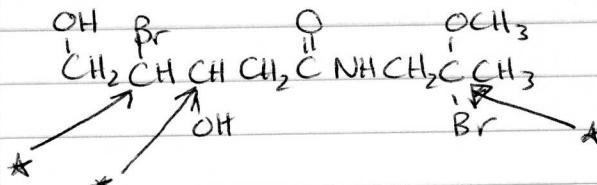
d.



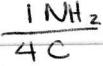
f.



3. Chiral C's - attached to 4 different substituents

4. most soluble (b) lots of OH groups, $\frac{3\text{OH}}{5\text{C}}$ can form lots of H-bonds to water.2nd (a) 3-C carboxylic acid - can H-bond to water $\frac{1\text{OH}}{3\text{C}}$

3rd (d) can H-bond to water but has a longer nonpolar part than a.



4th (c) ester - polar. can accept H-bonds, but no H-bond donating groups - not as soluble as (a)

(hard to decide between d and c which would be least soluble).

5. All of these have very similar molar masses, so similar London forces.

1. (a) highest bp - can form H-bonds to each other, very polar, can form "dimers" (2 stuck together)

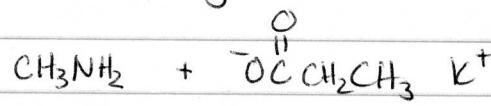
2. (d) can form H-bonds to each other, but not as strongly as a.

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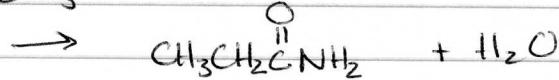
3. c. polar but can't H-bond to each other, so weaker IMF's than d.

b. totally nonpolar - only London forces - weakest IMF's overall.

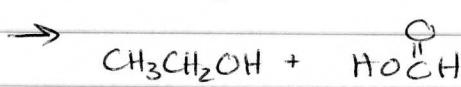
6. a. base hydrolysis of amide. Products:



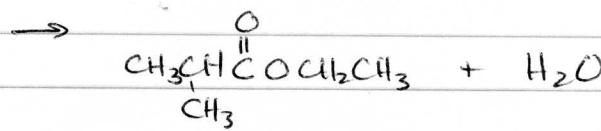
b. amidation



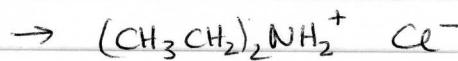
c. ester hydrolysis



d. esterification



e. amine + SA



7. amines: weak bases, smell bad (fishy, decay, etc)

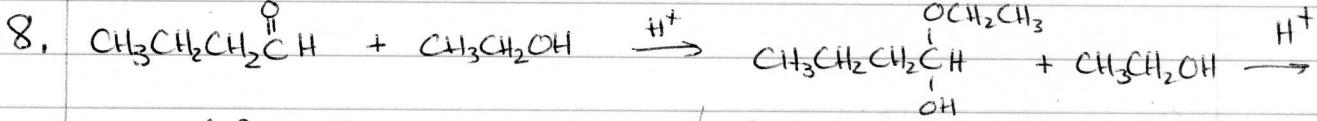
aldehydes - some smell good

ketones - excellent solvents

CA's - weak acids

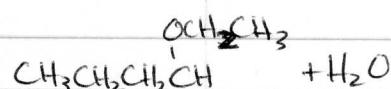
esters - smell good (fruit flavors)

amides - not acidic or basic

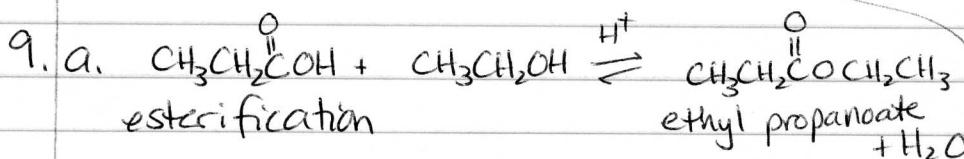


acetal formation

hemiacetal

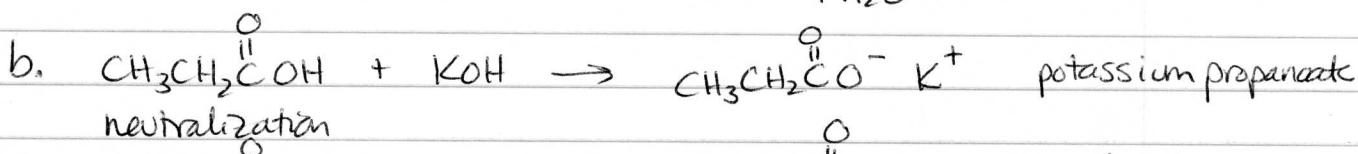


acetal

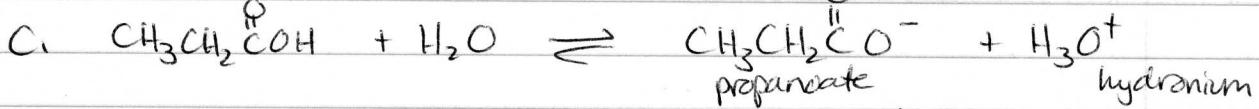


esterification

ethyl propanoate

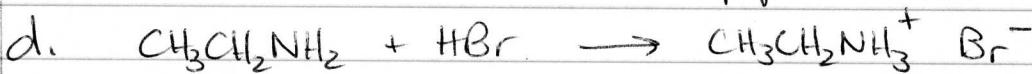


neutralization



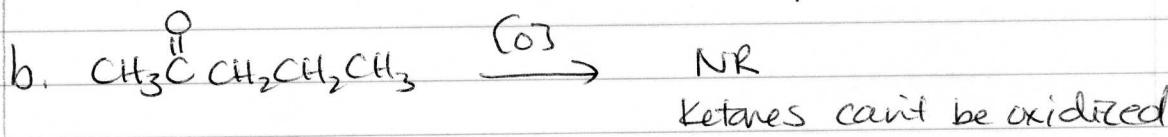
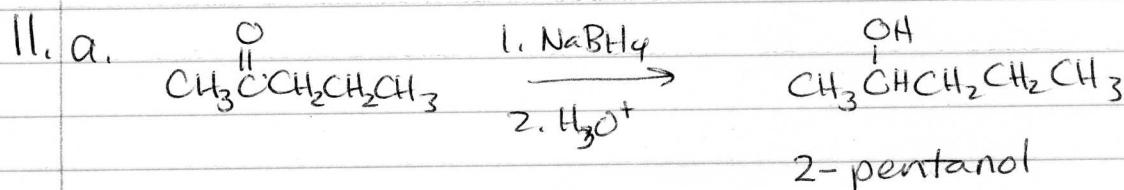
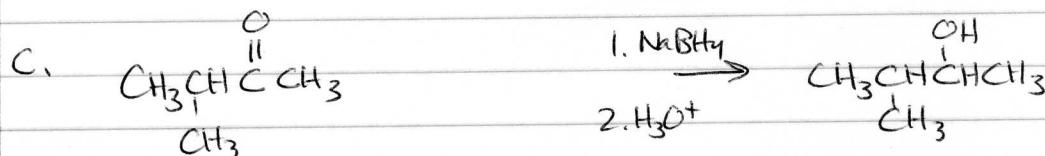
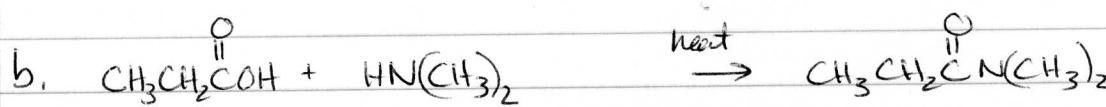
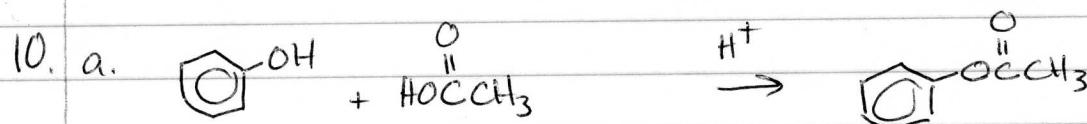
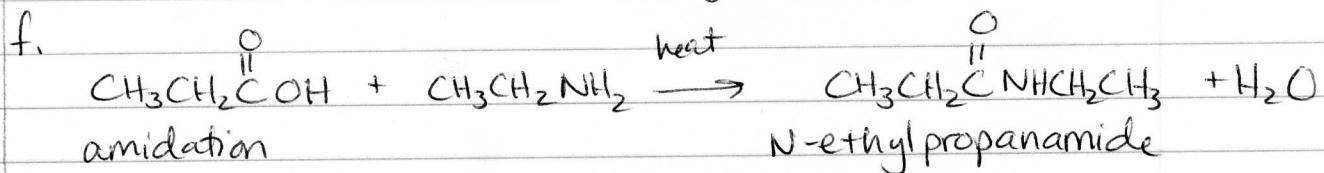
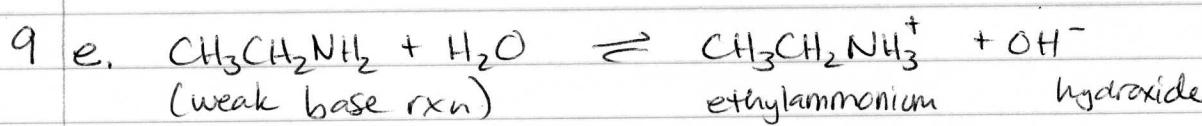
propanoate

hydronium



ethylammonium bromide

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12. a. Benedict's test

Cu^{2+} in basic solution \oplus - red ppt

if positive, the starting compound got oxidized.

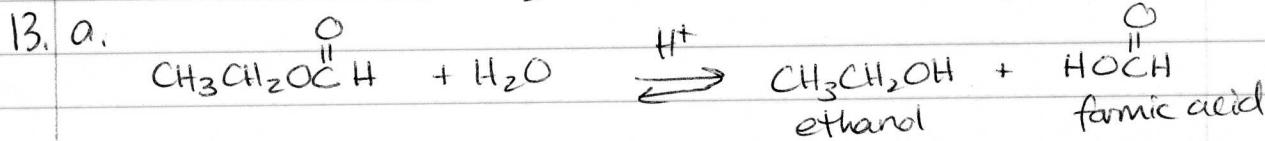
(it could be an alcohol or an aldehyde, but not a ketone)

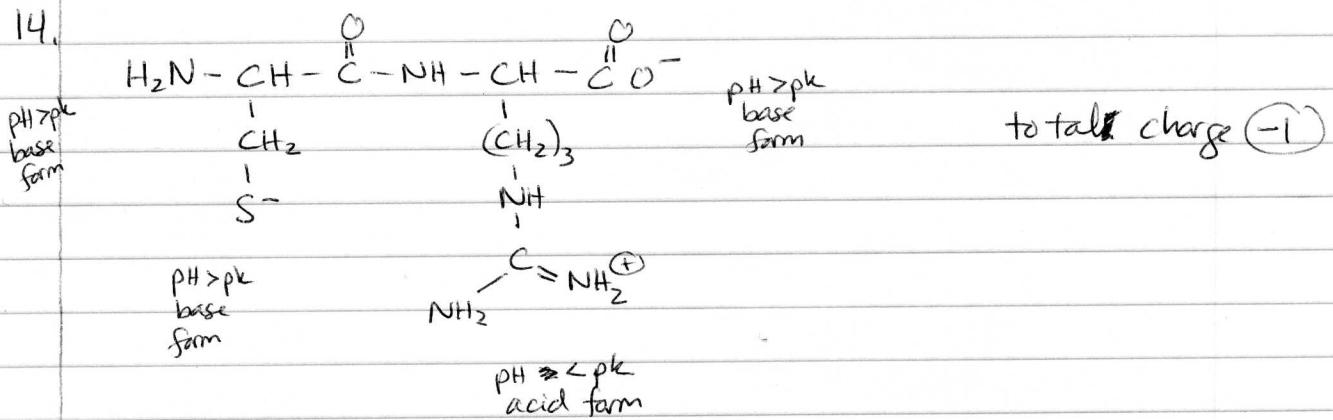
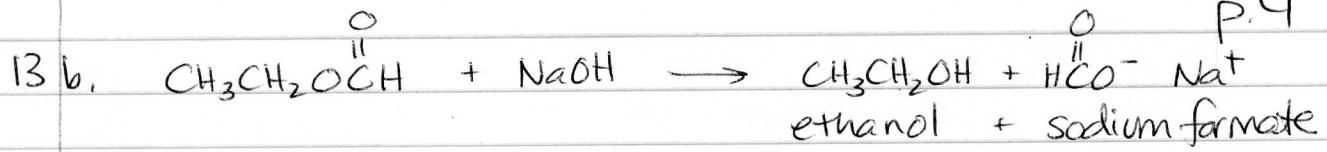
b. I_2 test - deep blue if positive.

if \oplus , starch is present.

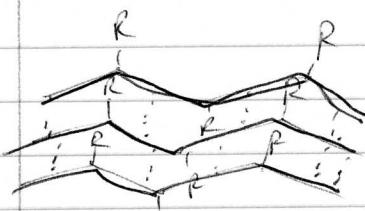
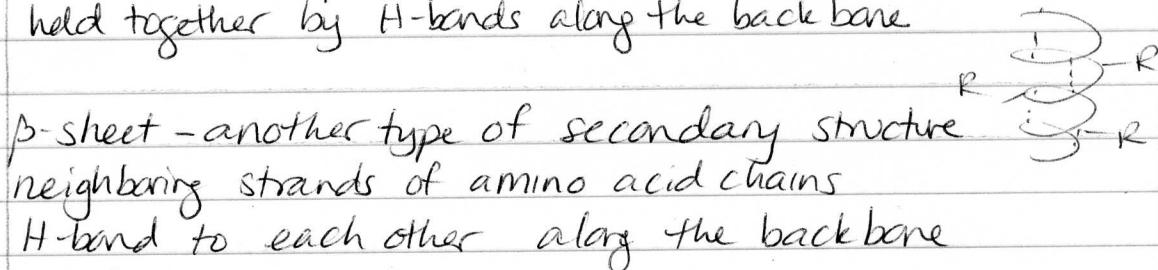
c. Br_2 test - if \oplus , orange color of Br_2 fades.

if \oplus , the compound has double or triple bonds that can react with Br_2 (it adds to double bond)





15. α -helix - a type of secondary structure - shape - helix or corkscrew.
 held together by H-bonds along the back bone



16. acidic conditions (or certain enzymes)
complete hydrolysis yields amino acids.
incomplete hydrolysis yields smaller peptide chains (than before)

- | | | | |
|--------------------------------|------------------|--|--|
| 17. a. Asp O^- | Arg N^+ | Salt bridge
+ and - charges | b. Gln, Asn
both have H-bond donating and accepting groups. |
| b. Leu and Ile - both nonpolar | | d. Cys, Cys - disulfide bridges
$-\text{CH}_2-\text{S}-\text{S}-\text{CH}_2-$ | |
| e. Thr, Lys polar | N^+ | H-bonding
both can donate or accept H-bonds | |

18. 1° Structure - sequence of aa's (held together by peptide bonds)
2° Structure - regular, repeating structures such as α -helix and β sheet. Held together by H-bonds between C=O and N-H groups along the backbone of the amino acid chain.
3° Structure - overall folding of the entire protein. Stabilized by interactions among the R groups: H-bonding, hydrophobic interactions, disulfide bridges, salt bridges, hydrophilic interactions.
(Also involves interaction of side chains with the solvent (water))
19. Denaturation - disruption of normal folding of protein ~~unfolding~~. When a protein is denatured, it can no longer function.
20. Normally, a protein folds so that the nonpolar side chains are on the inside of the protein, away from the water/solvent. Polar and charged groups are on the outside, facing the water/solvent. (There are also some polar and charged groups on the inside, interacting with each other.) In the presence of an organic solvent, the protein would fold differently. The nonpolar side chains will point toward the outside, facing the solvent (because nonpolar substances are compatible). The polar and charged groups would face inside to be away from the nonpolar solvent. The protein would basically turn inside out and be completely inactivated.
21. If strong acid is added, it causes all acidic groups, which are normally \ominus charged at pH 7, to become protonated and thus uncharged. This would disrupt all of the salt bridges that are stabilizing the 3° structure. (\oplus and \ominus charges attract, but \oplus and \oplus charges don't!) Protein will unfold partially and become inactivated.
22. Each enzyme has an active site that is specific and fits one particular substrate. Its catalytic groups are positioned to help catalyze one very specific reaction. Other substrates won't fit and won't have the appropriate catalytic groups available.

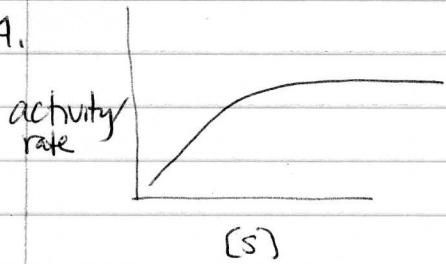
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23. The enzyme active site is complementary to the structure of the substrate, so it only fits one type of substrate. As the substrate binds, it is often forced into a more strained shape. This destabilizes the substrate (so it has higher energy) and helps lower the activation energy of the reaction.

Groups in the enzyme active site act as catalytic groups, providing whatever is needed for the reaction - (transfer protons, transfer e^- , stabilize charge, break/form new bonds, etc). The catalytic groups are positioned perfectly relative to the substrate in the active site - they give it exactly what it needs to react.

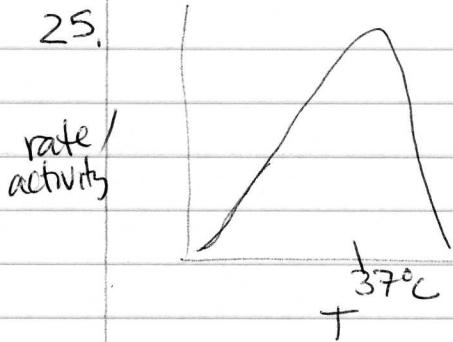
Rxn occurs, and product detaches from the enzyme.

24.



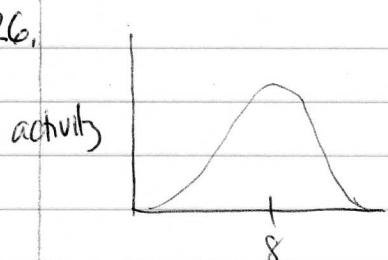
as $[S]$ increases, the rate increases, since there's a greater probability that E and S will collide. After enough S has been added to bind a S to every E, the E active sites are saturated. The enzyme keeps working, but adding more S won't help to speed it up. The E molecules are working as fast as they can.

25.



All reactions are faster at a higher temp, because more molecules will have enough energy to react. As temp keeps increasing, the enzyme becomes denatured, and can't function any more, so rate of rxn slows or stops.

26.



at low pH - very acidic solution - some side chains get protonated which changes $(-)$ charges to (\oplus) charges. This disrupts salt bridges and causes denaturation of the protein. a denatured protein can no longer function.

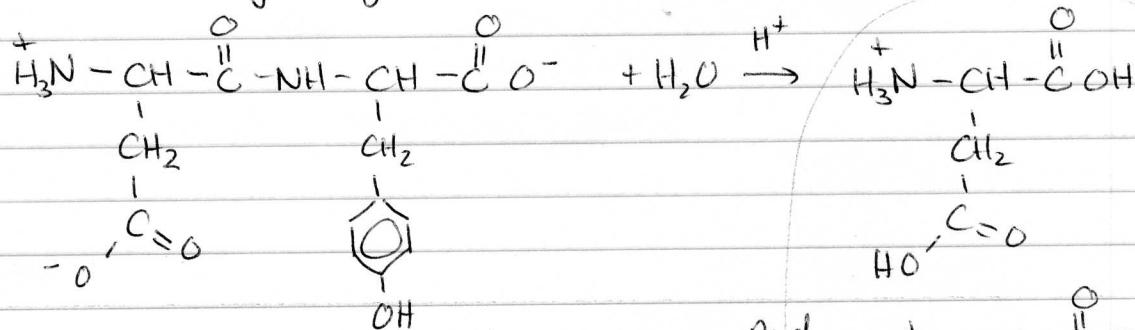
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27. a. substrate loses H⁺ being oxidized.
 b. this is a hydrolysis Rxn
 c. forming larger molecules, requiring energy
 d. NH₂ group and OH group are
 switched/transferred between molecules
- oxidoreductase
hydrolase
ligase
transferase

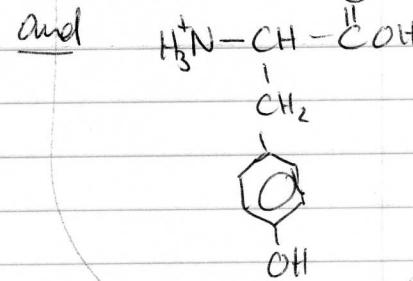
28. a cofactor is a non-protein part that's necessary for the protein to function.

29. as T[°], enzyme gets denatured.

30.



pH 7



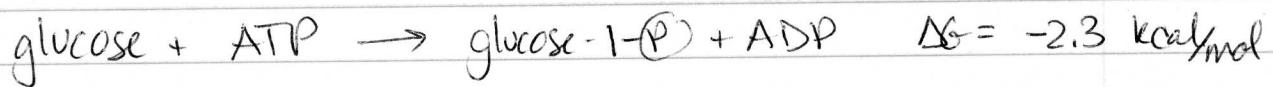
all groups
switched

31. Lock and key - substrate fits into enzyme active site like a key into a lock with no changing of shape, of either S or E.

Induced-fit - S or E or both change shape slightly as the substrate binds.

32. lots of N, NH, O, OH - this is polar and would be able to H-bond to water in lots of locations - so water soluble.

33. glucose + (P) → glucose-1-(P) + H₂O 5.0 kcal/mol
 ATP + H₂O → ADP + (P) -7.3 kcal/mol



ΔG is \ominus so spontaneous overall.

34. Competitive inhibitor - similar in structure to substrate.

This type of inhibitor will bind to the active site, but won't be able to react. When it's bound, the normal substrate can't bind. This will slow the rxn.

(At any time, some enzymes will bind inhibitor and some will bind substrate.)

If you add lots of excess substrate, enzyme activity can be restored.

35. A noncompetitive inhibitor binds to the enzyme somewhere other than the active site. It changes the shape of the enzyme, making the active site inaccessible so substrate can't bind. This will slow/stop the rxn.
(adding more substrate won't help.)

